

TWO NEW DITERPENES WITH A ROSANE SKELETON FROM VELLOZIACEAE.

ANGELO C. PINTO^a, TEREZA CRISTINA V. SCOFIELD^a, and RAIMUNDO BRAZ FILHO^b.

^aNúcleo de Pesquisas de Produtos Naturais, Universidade Federal do Rio de Janeiro, CCS, Bloco H, 21941, Rio de Janeiro, RJ, Brazil. ^bDepartamento de Química, Universidade Federal Rural do Rio de Janeiro.

SUMMARY: Two new rosane diterpenes, epoxicorcovadin and epoxivellozin, have been isolated from Vellozia candida Mikán: their structures were established by spectroscopic data and chemical transformations.

Vellozia candida Mikán grows on the hillside of Corcovado mountain and on the rocky mountains of the coast of State of Rio de Janeiro. We have previously reported the isolation of velloziolide (1)¹ and of three rosane diterpenes² from the hexane extract of the stem, roots and leaf sheaths of this plant. The present paper describes the structural elucidation of two new diterpenoids, epoxicorcovadin (2a) and epoxivellozin (3a), isolated as minor components from the same plant.

Epoxicorcovadin (2a) was purified and characterized as its acetate derivative (2b). 2b [C₂₂H₃₀O₄; m.p. 168-69°C, [α]_D²⁵ = 0.00 (c = 1.00 CHCl₃)]³ possesses as a characteristic feature an α,3-unsaturated ketone moiety (ν_C = 0 at 1668 cm⁻¹ and λ_{max}^{EtOH} at 253 nm). The ¹H NMR spectrum, in CDCl₃, showed signals for four tertiary methyl groups (0.98, 1.10, 1.18 and 1.26 ppm), a methyl group of an acetate (2.09 ppm) and one tertiary vinyl group at 5.67 ppm (1H, dd, J=17 and 10 Hz). The multiplet signal (3H) between at 4.98 and 5.31 ppm (CH₂, m) corresponds to the methylene of the vinyl residue and the acetyl carbinolic proton. Two doublets at 5.9 (1H, J=9 Hz) and 7.45 ppm (1H, J=9 Hz) are due to the two olefinic protons α,3-conjugated to the carbonyl group, with the 3 carbon linked to a quaternary carbon. Double irradiation experiments demonstrated that these two protons are mutually spin coupled. The absence of absorptions in the IR spectrum for hydroxyl groups and the presence of two singlet signals at 64.3 and 68.4 ppm in ¹³C NMR spectrum of 2b indicated that the remaining oxygen atom belongs to an epoxide function. These data, in combination with the comparative analysis of the proton noise decoupled and single frequency off-resonance decoupled ¹³C NMR spectrum (Table 1), allowed expansion of the molecular formula C₂₂H₃₀O₄ to (CO-CH=CH-■)(CHOAc)(■-CH=CH₂)(■-C^O-C-■)(C)₃(CH)(CH₂)₄(CH₃)₄ and to deduce the structure 2b, which was confirmed by chemical

transformation. Thus, epoxidation of 4² with m-CPBA at 10°C, in CH₂Cl₂, afforded the 5 α ,10 α -epoxide 5 as the main product by attack of the bulky reagent from the less hindered α -face. Stereochemical assignment of the epoxy ring of 5 can be made by comparative ¹³C NMR analysis of 2 and 4. Thus in 2b, C-8 appears at a more shielded position (32.7 ppm) than in 4 (37.7 ppm, Table 1). This can be explained by a shielding effect due to the "γ-gauche like" relationship between the C-8-H and the 5 α ,10 α -oxirane ring in 2. Treatment of epoxide 5 with LDA and phenylselenyl bromide⁴ in dry THF at -78°C followed by addition of hydrogen peroxide at room temp, furnished a product that upon acetylation was identical in all aspects with natural 2.

Epoxivellozin [3a; C₂₀H₂₈O₄; m.p. 142-4°C; [α]_D²⁵ = + 14.9 (c= 0.77 CHCl₃)] revealed in the IR spectrum the presence of a conjugated ketone (1630 cm⁻¹, broad band), hydroxyl (3420cm⁻¹, broad band), and vinyl (900 cm⁻¹) groups. The UV spectrum presented bands at $\lambda_{\max}^{\text{EtOH}}$ 228 and 291 nm, which were shifted to $\lambda_{\max}^{\text{EtOH}}$ 229 and 338 nm after addition of one drop a NaOH solution, indicating the presence of a diosphenol chromophore.⁵

Table 2 lists the ¹H NMR spectral data of 3b assigned to the corresponding protons by integration, multiplicity and nuclear magnetic double resonance techniques. The smaller value of the coupling constant of one of the protons at C-14 (J= 2.5Hz) indicates its equatorial position (W-type coupling with Heq-12). The presence of two hydroxyl group was confirmed by acetylation of 3a (Ac₂O/DMAP) to yield a diacetyl derivative 3b showing two methyl singlets at 2.02 and 2.24 ppm (acetyl groups) and a double doublet at 5.24 ppm which corresponds to a carbinolic proton deshielded by acetylation (4.04 ppm in 3). The UV spectrum of 3b showed $\lambda_{\max}^{\text{EtOH}}$ at 252 nm. The hypsochromic shift observed in the conversion of 3a to 3b confirmed the presence of the diosphenol chromophore in 3a.⁵

The remaining oxygen atom in 3a was assigned to a tetrasubstituted epoxide moiety on the basis of ¹³C NMR peaks observed at 66.9 (s) and 72.3 (s) ppm. Reduction of 3b with NaBH₄ in MeOH at room temp gave monoacetate (3c) ($\lambda_{\max}^{\text{EtOH}}$ 228 and 291 nm). The ¹³C NMR spectral data of 3a and 3b (Table 2) in combination with biogenetic considerations allowed postulation of structure 3a for epoxivellozin.

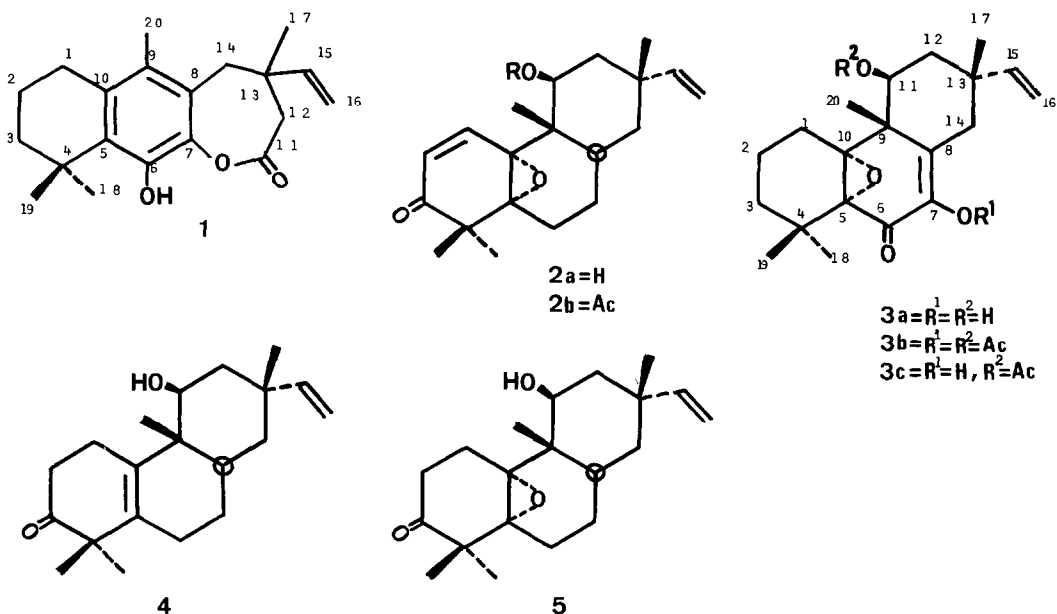
The fact that compound 3a occurs in the same plant with velloziolide 1 allows us to consider 3a as a possible biogenetic precursor for the former diterpenoid.

Table 1. ^{13}C n.m.r. spectral data (δ) for compounds 2b, 3a, 3b, 4 and 5, (TMS as internal standard, in CDCl_3)

Carbon	<u>2b</u>	<u>3a</u>	<u>3b</u>	<u>4</u>	<u>5</u>	
1	145.4	28.1	28.2	25.1	22.4	
2	128.4	17.9	17.7	37.1	33.7	
3	201.0	37.7	36.7	216.0	213.0	
4	48.2	31.8	31.9	47.6	48.9	
5	64.3	72.3	70.4	133.1	66.9	
6	23.0	189.7	186.9	24.9	22.4	
7	23.5	140.2	138.7	27.4	23.1	
8	32.7	128.9	143.8	37.7	31.8	
9	39.9	45.3	45.4	42.9	42.0	
10	68.4	66.9	68.5	136.4	68.9	
11	74.7	70.9	72.5	72.9	70.8	
12	40.2	44.0	39.0	43.6	43.4	
13	38.0	38.0	37.5	37.3	37.4	
14	37.0	33.3	33.3	40.5	40.1	
15	146.5	143.6	142.5	146.6	145.9	
16	113.5	112.9	113.9	112.1	112.3	
17	31.1	30.0	29.9	31.1	30.9	
18	17.3	23.9	23.4	25.8	18.7	
19	24.3	26.2	26.0	23.0	22.4	
20	11.2	17.3	19.3	11.6	8.9	
COO	169.7	-	169.1	167.8	-	..
CH ₃	21.6	-	20.2	21.5	-	..

Table 2. ^1H n.m.r. spectral data (δ) including decoupling experiments for vellozioepoxi 3b (400MHz, TMS as internal standard, in CDCl_3)

Heq-1:	2.60 (dt, J= 15 and 4Hz)	
Hax-1:	1.97 (ddd, J= 15,11 and 5Hz)	
2H-2	1.70-1.20	
2H-3		
Hax-11:	4.05 (dd, J= 11 and 4.4Hz)	←
Heq-12:	1.78 (ddd, J= 13, 4.4 and 2.5Hz)	←
Hax-12:	1.68 (dd, J= 13 and 11Hz)	←
Heq-14:	3.02 (dd, J= 15 and 2.5Hz)	←
Hax-14:	1.92 (d, J= 15Hz)	←
H-15:	5.60 (dd, J= 18.0 and 11.3Hz)	
H-16:	5.02 (d, J= 18Hz)	
H-16:	4.98 (d, J= 11.3Hz)	
CH ₃ -17	1.09 (s)	
CH ₃ -18	1.30 (s)	
CH ₃ -19	1.22 (s)	
CH ₃ -20	1.34 (s)	



We are grateful to Profa. Nanuza L. de Menezes (Department of Botany, USP, Brazil) for the identification of *Vellozia candida* Mikan, SARSA for optical rotation data and Prof. Gabor Lukacs (Gif-sur-Yvette) for the 400MHz ¹H NMR spectrum of epoxivellozin 3.

1. A.C. Pinto, M.L.A. Gonçalves, R. Braz Fç, A. Neszmelyi and G. Lukacs, J. Chem. Soc. Chem. Commun., 293, 1982.
2. A.C. Pinto, W.S. Garcez, M.L.G. Ficara, T.C. Vasconcelos, A.L. Pereira, L.N.L.F. Gomes, M.C. Frechiani and M.L. Patitucci, An. Acad. brasil. Ciênc., 54, 103 (1982).
3. $[\alpha]_{25}^{\lambda} = \frac{589}{0.00} \quad \frac{578}{0.002} \quad \frac{546}{0.011} \quad \frac{436}{0.21} \quad \frac{365}{1.69} \quad (c = 1,0 \text{ CHCl}_3)$
4. P.A. Grieco and M. Myashita, J. Org. Chem., 39, 120 (1974).
5. A.I. Scott, "Interpretation of the Ultraviolet Spectra of Natural Products", Pergamon Press, Oxford, New York, p.60 (1974).

(Received in USA 19 August 1983)